### Media Recipes for Ribes

#### Ribes growth medium (solid) - 1000 ml

✓ To a small volume of double distilled water (ddH₂0) add:

Potassium nitrate (KNO <sub>3</sub> )	0.576 g
Ammonium nitrate (NH <sub>4</sub> KNO <sub>3</sub> )	0.5 g
Magnesium sulfate (MgSO <sub>4</sub> )	0.1807 g
Calcium chloride, dihydrate (CaCl <sub>2</sub> )	0.332 g
Potassium phosphate, <i>monobasic</i> (KH <sub>2</sub> PO <sub>4</sub> )	0.17 g
Iron stock <sup>1</sup>	20.0 ml
MS micronutrients <sup>1,2</sup>	10.0 ml
MS vitamins <sup>1,2</sup>	10.0 ml
Ascorbic acid	0.5 g
Glucose	20.0 g
GA <sub>3</sub> (gibberellic acid)	0.2 mg
BA (6-benzylaminopurine)	0.1 mg

- ✓ Stir until well blended
- ✓ Bring to final volume (1000 ml) with ddH<sub>2</sub>0
- ✓ Adjust pH to 5.7
- ✓ Add:

Gellan gum (Phytagel™³ˆ)	3.5 g
Agar (Sigma <sup>®3</sup> A7002*)	1.45 g

- ✓ Heat and stir until boiling
- ✓ Dispense into stacked Magenta®⁴ GA7\* culture vessels (40 ml/vessel)
- ✓ Autoclave

## Iron stock solution (100x) (liquid) - 500 ml

✓ To a small volume of double distilled water (ddH₂0) add:

NaEDTA, disodium salt, dihydrate (NA2EDTA\*2H20) 1.86 g

- ✓ Stir until NaEDTA is completely dissolved
- ✓ In a separate vessel containing a small volume of ddH<sub>2</sub>0 add:

Ferric sulfate (FeSO<sub>4\*</sub>7H<sub>2</sub>O) 1.39 g

- ✓ Heat and stir until the ferric sulfate is completely dissolved. Allow solution to cool completely
- ✓ Combine the NaEDTA solution with the ferric sulfate solution

- ✓ Bring to volume (500 ml) and stir until the solution turns yellow
- ✓ Dispense into an amber vessel to prevent photodegredation. Store at 2-4 °C

## MS<sup>3</sup> micronutrient stock solution (100x) (liquid) – 500 ml

✓ To a small volume of double distilled water (ddH₂0) add:

Boric acid (H <sub>3</sub> BO <sub>3</sub> )	0.31 g
Cobalt chloride (CoCl <sub>2</sub> * 6H <sub>2</sub> 0)	0.00125 g
Cupric sulfate (CuSO <sub>4*</sub> 5H <sub>2</sub> 0)	0.00125 g
Zinc sulfate (ZnSO <sub>4*</sub> 7H <sub>2</sub> 0)	0.43 g
Molybdic acid, sodium salt, dihydrate (NaMoO <sub>4*</sub> 2H <sub>2</sub> O)	0.0125 g
Manganese sulfate (MnSO <sub>4*</sub> H <sub>2</sub> 0)	0.845 g
Potassium iodide (KI)	0.0415 g

- ✓ Heat and stir until boiling and dry ingredients have completely dissolved
- ✓ Bring to final volume (500 ml) with ddH<sub>2</sub>0
- ✓ Dispense into desired vessel and store at 2-4 °C or aliquot and store in freezer

# MS<sup>3</sup> vitamin stock solution (100x) (liquid) – 500 ml

✓ To a small volume of double distilled water (dd $H_2$ 0) add:

Glycine (free base)	$0.1\mathrm{g}$
Myo-inositol	5.0 g
Nicotinic acid (free base)	0.025 g
Pyridoxine HCl	0.025 g
Thiamine HCl	0.005 g

- ✓ Stir until ingredients are well blended
- ✓ Bring to final volume (500 ml) with ddH20
- ✓ Dispense into desired vessel and store at 2-4 °C or aliquot and store in freezer

## Ca-free MS+3% (w/v) Na-alginate medium (liquid) – 100 ml

✓ To a small volume of double distilled water (dd $H_2$ 0) add:

Potassium nitrate (KNO <sub>3</sub> )	0.19 g
Ammonium nitrate (NH <sub>4</sub> KNO <sub>3</sub> )	0.165 g
Magnesium sulfate (MgSO <sub>4</sub> )	0.018 g
Potassium phosphate, monobasic (KH <sub>2</sub> PO <sub>4</sub> )	0.017 g
Iron stock <sup>1</sup>	1.0 ml

MS micronutrients<sup>1,2</sup>

MS vitamins<sup>1,2</sup>

Reagent grade sucrose

1.0 ml

25.67 g

- ✓ Stir until sucrose is completely dissolved
- ✓ Bring to final volume (100 ml) with ddH₂0
- ✓ Adjust pH to 5.8
- ✓ Add:

Alginic acid sodium salt, 2% viscosity (Sigma<sup>®3</sup> A2158\*) 3.0 g

To prevent clumping, add the alginic acid slowly to rapidly stirring medium.

A homogenizer with a propeller-type stirring blade works well for this step.

- ✓ Continue stirring until well blended and alginic acid is completely dissolved (~20 minutes)
- ✓ Dispense into desired vessels
- ✓ Autoclave

#### 100 mM calcium chloride+MS encapsulation medium (liquid) – 1000 ml

 $\checkmark$  To a small volume of double distilled water (ddH<sub>2</sub>0) add:

MS basal medium w/vitamins<sup>2</sup> 4.43 g (prepackaged as M519<sup>5</sup>) Calcium chloride, dihydrate (CaCl<sub>2</sub>) 14.7 g Sucrose 30.0 g

- ✓ Stir until dry ingredients are completely dissolved
- ✓ Bring to final volume (1000 ml) with ddH₂0
- ✓ Adjust pH to 5.7
- ✓ Heat and stir until well blended
- ✓ Dispense into desired vessels
- ✓ Autoclave

#### 0.75 M sucrose+MS medium (liquid) - 1000 ml

✓ To a small volume of double distilled water (ddH₂0) add:

MS basal medium w/vitamins<sup>2</sup> 4.43 g (prepackaged as M519<sup>5</sup>) Sucrose 256.72 g

✓ Stir until sucrose is completely dissolved

- ✓ Bring to final volume (1000 ml) with ddH<sub>2</sub>0
- ✓ Adjust pH to 5.7
- ✓ Mix and heat until boiling
- ✓ Dispense into desired vessels
- ✓ Autoclave

#### MS rehydration medium (liquid) – 1000 ml

✓ To a small volume of double distilled water (ddH₂0) add:

MS basal medium w/vitamins<sup>1</sup> 4.43 g (prepackaged as M519<sup>5</sup>) Sucrose 30.0 g

- ✓ Stir until dry ingredients are completely dissolved
- ✓ Bring to final volume (1000 ml) with ddH₂0
- ✓ Adjust pH to 5.7
- ✓ Mix and heat until boiling
- ✓ Dispense into desired vessels
- ✓ Autoclave

## Ribes Ca-free MS medium (liquid) - 1000 ml

✓ To a small volume of double distilled water ( $ddH_2O$ ) add:

Potassium nitrate (KNO <sub>3</sub> )	0.576 g
Ammonium nitrate (NH <sub>4</sub> KNO <sub>3</sub> )	0.5 g
Magnesium sulfate (MgSO <sub>4</sub> )	0.1807 g
Potassium phosphate, monobasic (KH <sub>2</sub> PO <sub>4</sub> )	0.17 g
Iron stock <sup>1</sup>	20.0 ml
MS micronutrients <sup>1,2</sup>	10.0 ml
MS vitamins <sup>1,2</sup>	10.0 ml
Ascorbic acid	0.5 g
Glucose	20.0 g

- ✓ Stir until dry ingredients are completely dissolved
- ✓ Bring to final volume (1000 ml) with ddH<sub>2</sub>0
- ✓ Adjust pH to 5.7
- ✓ Dispense into desired vessels

#### ✓ Autoclave

## Ribes recovery medium with PVP (solid) - 1000 ml

✓ To a small volume of double distilled water (ddH₂0) add:

0.576 g
0.5 g
0.1807 g
0.332 g
0.17 g
20.0 ml
10.0 ml
10.0 ml
0.5 g
20.0 g
0.2 mg
0.1 mg
1.0 g

- ✓ Stir until dry ingredients are completely dissolved
- ✓ Bring to final volume (1000 ml) with ddH<sub>2</sub>0
- ✓ Adjust pH to 5.7
- ✓ Add:

Gellan gum (Phytagel™³)	3.5 g
Agar (Sigma <sup>®3</sup> A7002*)	1.45 g

- ✓ Heat and stir until boiling
- ✓ Autoclave
- ✓ In laminar flow hood, dispense slightly cooled liquid into Petri dishes
- ✓ <sup>1</sup> Recipe follows
- ✓ <sup>2</sup> Murashige & Skoog, 1962
- ✓ <sup>3</sup> Sigma-Aldrich, St. Louis, MO\*
- √ <sup>4</sup> Magenta Corp., Chicago, IL\*
- √ <sup>5</sup>Phytotechnology Laboratories, Shawnee Mission, KS\*

<sup>\*</sup> Mention of trade names or commercial products in this article is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture.